



Isolation and structures of hedathiosulfonic acids A and B, novel thiosulfonic acids from the deep-sea urchin *Echinocardium cordatum*

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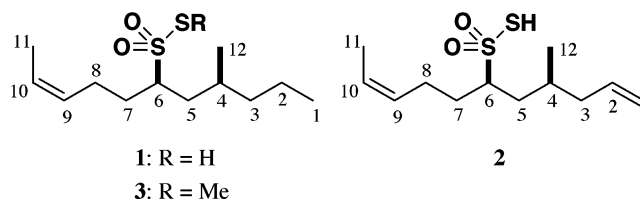
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Abstract—Hedathiosulfonic acids A and B were isolated from the deep-sea urchin *Echinocardium cordatum*, and were determined to be novel 6-undecanethiosulfonic acids by 2D NMR, HRMS, and methylation reaction. Hedathiosulfonic acids exhibited acute toxicity. © 2001 Elsevier Science Ltd. All rights reserved.

In our continuing search for marine bioactive compounds, we have reported the isolation and structural determination of pinnatoxins, halichlorine, haterumalides and pinnamine.¹ To discover new bioactive compounds, we have directed our attention to the deep-sea invertebrates as sources for such compounds. As a result, we have recently isolated hedathiosulfonic acids, which have a thiosulfonic acid functionality. We report here the isolation and structure determination of hedathiosulfonic acids A (**1**) and B (**2**) from the deposit-feeding deep-sea heart urchin *Echinocardium cordatum*.

The aqueous 80% EtOH extract of the deep-sea urchin *E. cordatum*, collected off the Heda coast of the Izu Peninsula, was partitioned between EtOAc and H₂O. The EtOAc-soluble fraction was subjected to fractionation guided by acute toxicity against mice using column chromatography (ODS, aqueous 60% MeOH) and reversed-phase HPLC (ODS, 50% MeOH) to give hedathiosulfonic acids A (**1**, 0.0039% yield based on wet wt) and B (**2**, 0.0028% yield based on wet wt) as colorless oils.² Hedathiosulfonic acids A (**1**) and B (**2**) exhibited low acute toxicity against mice, with LD_{99s} of 0.39 and 0.36 g/kg, respectively.



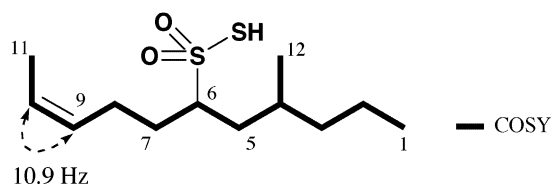
The fluorescent X-ray analysis of hedathiosulfonic acid A (**1**) suggested the presence of two sulfur atoms. The molecular formula of **1** was determined to be C₁₂H₂₄O₂S₂ by HRFABMS (*m/z* 309.0951, calcd for C₁₂H₂₃NaO₂S₂, 309.0935). The NMR data for **1** are summarized in Table. The ¹H NMR, ¹³C NMR, and HMQC spectra of **1** showed the presence of three methyl carbons, five methylene carbons, two methine carbons, and two olefinic methine carbons (δ_C 125.3, 131.3 ppm). Based on the chemical shift, it was clarified that one methine carbon was attached to an oxygenated sulfur atom (δ_H 2.86 ppm, δ_C 70.8 ppm). A detailed analysis of COSY spectrum enabled us to elucidate the entire carbon framework (Fig. 1). Furthermore, to determine the functional group (S₂O₂H) at C-6, hedathiosulfonic acid A (**1**) was methylated with Me₂SO₄–Et₃N to give methyl thiosulfonate **3**. The chemical shifts of an additional methyl group (δ_H 2.67 ppm, δ_C 17.5 ppm) in **3** suggested that this methyl carbon was attached to a non-oxygenated sulfur atom. As a result, we determined that hedathiosulfonic acid A (**1**) had a thiosulfonic acid functionality. Finally, the geometry of the C-9 olefin was determined to be *9Z*.

Keywords: *Echinocardium cordatum*; natural thiosulfonic acid; isolation; structure determination; acute toxicity.

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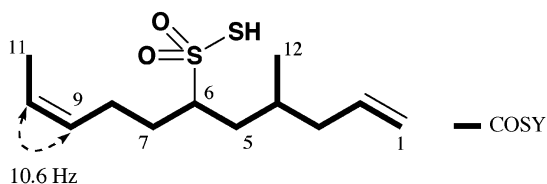
Table 1. NMR data for hedathiosulfonic acid **1** and **2** in CD₃OD

Hedathiosulfonic acid 1			Hedathiosulfonic acid 2		
Atom	¹ H ^a	¹³ C ^b	Atom	¹ H ^a	¹³ C ^b
1	0.91 t 3H (7.2)	14.7 q	1a	4.98 br d (17.4)	116.2 t
2a	1.32 m	20.9 t	1b	5.01 br d (10.1)	
2b	1.44 m		2	5.83 ddt (10.1, 17.4, 7.2)	138.4 d
3a	1.11 m	40.1 t	3a	1.88 m	
3b	1.39 m		3b	2.19 m	42.1 t
4	1.78 m	31.8 d	4	1.88 m	32.1 d
5a	1.31 m	39.9 t	5a	1.34 ddd (6.3, 6.6, 14.5)	39.5 t
5b	2.11 ddd (5.3, 7.3, 14.3)		5b	2.13 ddd (5.8, 6.7, 14.5)	
6	2.86 dq (5.3, 5.1)	70.8 d	6	2.88 ddt (6.3, 5.8, 5.9)	70.7 d
7a	1.58 m	32.7 t	7a	1.57 m	32.7 t
7b	2.18 m		7b	2.19 m	
8a	2.23 ddt (7.7, 14.5, 7.2)	25.9 t	8a	2.23 ddt (7.2, 14.6, 7.6)	25.8 t
8b	2.30 ddt (8.1, 14.5, 8.0)		8b	2.29 m	
9	5.40 br ddd (7.7, 8.1, 10.9)	131.3 d	9	5.40 dddq (6.7, 7.2, 10.6, 1.7)	131.2 d
10	5.47 br dq (10.9, 6.7)	125.3 d	10	5.47 br dq (10.6, 6.7)	125.3 d
11	1.63 d 3H (6.7)	13.0 q	11	1.63 br d 3H (6.7)	13.1 q
12	0.92 d 3H (6.6)	20.6 q	12	0.93 d 3H (6.2)	20.2 q

^a Recorded at 800 MHz. Coupling constants (Hz) are in parentheses.^b Recorded at 200 MHz. Multiplicity was based on HMQC spectrum.**Figure 1.** Partial structures of hedathiosulfonic acid **1** based on 2D NMR correlations.

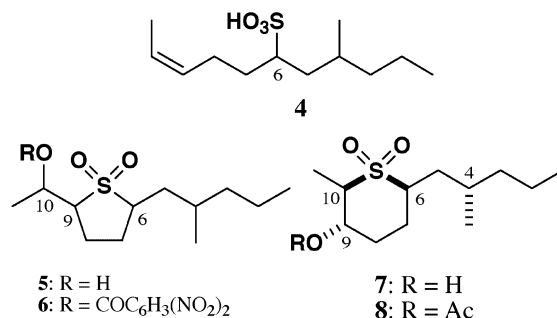
based on the coupling constant (10.9 Hz) between H-9 and H-10. Thus, the gross structure of hedathiosulfonic acid **1** was determined to be **1**.

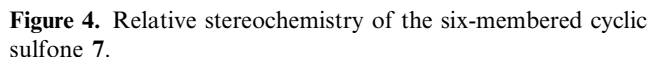
The ¹H NMR spectrum of hedathiosulfonic acid **2** resembled that of hedathiosulfonic acid **1**, but the presence of three additional vinyl protons (δ_{H} 4.98, 5.01, 5.83 ppm) and the molecular formula of **2** determined by HRFABMS (m/z 307.0761, calcd for C₁₂H₂₁Na₂O₂S₂, 307.0778) suggested that **2** was a dehydro derivative of **1**. The NMR data for **2** are summarized in Table 1. As expected, the gross structure of hedathiosulfonic acid **2** was determined to be **2** by similar analyses of COSY, HMQC, and HMBC spectra (Fig. 2).

**Figure 2.** Partial structures of hedathiosulfonic acid **2** based on 2D NMR correlations.

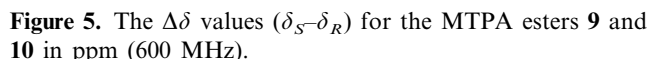
Hedathiosulfonic acid **1** was unstable and gradually decomposed during separation processes. Hedathiosulfonic acid **1** was transformed into sulfonic acid **4**, five-membered cyclic sulfone **5**, and six-membered cyclic sulfone **7** (Fig. 3).³ The gross structures of **4**, **5**, and **7** were determined using the 2D NMR and the mass spectra. Furthermore, to confirm the position of the hydroxyl group, sulfones **5** and **7** were converted into 3,5-dinitrobenzoate **6** and acetate **8**, respectively. Both the oxymethine protons H-10 in **6** and H-9 in **8** were shifted more than 1 ppm downfield from those in **5** and **7**.⁴

Because the stereochemistry in hedathiosulfonic acid **1** could not be deduced by the spectroscopic analysis, that of the sulfone **7** was determined as follows. As the six-membered sulfone ring part, the magnitude of $J_{8a,9} = 10.0$, $J_{8b,9} = 3.9$ and $J_{9,10} = 10.0$ Hz suggested that H-8a and H-9, H-9 and H-10 were located in *anti* arrangement, respectively. In addition, H-8b and H-9 were located in *gauche* arrangement (Fig. 4). In the NOE experiments (600 MHz) on **7**, irradiation of the signal at H-6 enhanced the signals for H-8a and H-10.

**Figure 3.** The degraded products of hedathiosulfonic acid **1** and those related compounds.



The absolute stereochemistry in **7** was determined using the modified Mosher's method (Fig. 5).⁵ Treatment of the sulfone **7** with (*R*)- or (*S*)-MTPACl gave (*S*)- or (*R*)-MTPA esters **9** and **10**, respectively. The ¹H NMR signals of the two MTPA esters **9** and **10** were assigned on the basis of the 2D NMR spectra, and the $\Delta\delta$ values ($\delta_S - \delta_R$, ppm) were then calculated. The results indicated that the absolute stereochemistry of C-9 was 9*S*, and that the absolute stereochemistry in **7** was suggested to be 4*S*, 6*R*, 9*S* and 10*R*. Therefore, the absolute stereochemistry in hedathiosulfonic acid A (**1**) was established to be 4*S* and 6*R*.



Acknowledgements

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References

- (a) Takada, N.; Sato, H.; Suenaga, K.; Arimoto, H.; Yamada, K.; Ueda, K.; Uemura, D. *Tetrahedron Lett.* **1999**, *40*, 6309; (b) Takada, N.; Umemura, N.; Suenaga, K.; Chou, T.; Nagatsu, A.; Haino, T.; Yamada, K.; Uemura, D. *Tetrahedron Lett.* **2001**, *42*, 3491; (c) Takada, N.; Iwatsuki, M.; Suenaga, K.; Uemura, D. *Tetrahedron Lett.* **2000**, *41*, 6425; (d) Kuramoto, M.; Chou, T.; Yamada, K.; Chiba, T.; Hayashi, T.; Uemura, D. *Tetrahedron Lett.* **1996**, *37*, 3867; (e) Arimoto, H.; Hayakawa, I.; Kuramoto, M.; Uemura, D. *Tetrahedron Lett.* **1998**, *39*, 861; (f) Chou, T.; Kuramoto, M.; Otani, Y.; Shikano, M.; Yazawa, K.; Uemura, D. *Tetrahedron Lett.* **1996**, *37*, 3871.
- Conditions for the isolation of hedathiosulfonic acid A (**1**): Develosil ODS HG-5 (ϕ 20×250 mm); solvent, 50%

- MeOH; flow rate, 5.0 mL/min; detection at 205 nm. $[\alpha]_D^{26} +2.1^\circ$ (c 0.073, MeOH); IR (CHCl₃) 3580–3260 (br), 1180, 1060 cm⁻¹. Conditions for the isolation of hedathiosulfonic acid B (**2**): Develosil ODS HG-5 (ϕ 20×250 mm); solvent, 50% MeOH; flow rate, 5.0 mL/min; detection at 205 nm. $[\alpha]_D^{26} -2.2^\circ$ (c 0.28, MeOH); IR (CHCl₃) 3520–3240 (br), 1640, 1180, 1060 cm⁻¹.
3. Studies on this reaction mechanism are in progress, see: Serra, A. C.; da Silva Corrêa, C. M. M. *Tetrahedron Lett.* **1991**, 32, 6653.
4. Compound **4**: ¹H NMR (CD₃OD) δ 2.72 (1H, H-6); FABMS m/z 293 (M-H+2Na)⁺. Compound **5**: ¹H NMR (CD₃OD) δ 4.07 (1H, H-10), 3.10 (1H, H-6), 2.90 (1H, H-9); FABMS m/z 271 (M+Na)⁺. Compound **6**: ¹H NMR (CD₃OD) δ 5.59 (1H, H-10), 3.44 (1H, H-9), 3.11 (1H, H-6); FABMS m/z 465 (M+Na)⁺. Compound **7**: ¹H NMR (CD₃OD) δ 3.51 (1H, H-9), 3.11 (1H, H-6), 2.93 (1H, H-10); FABMS m/z 271 (M+Na)⁺. Compound **8**: ¹H NMR (CD₃OD) δ 4.80 (1H, H-9), 3.30 (1H, H-10), 3.15 (1H, H-6); FABMS m/z 313 (M+Na)⁺.
5. Ohtani, I.; Kusumi, T.; Kashman, Y.; Kakisawa, H. *J. Am. Chem. Soc.* **1991**, 113, 4092.
6. (a) Turner, E.; Klevit, R.; Hager, L. J.; Shapiro, B. M. *Biochemistry* **1987**, 26, 4028; (b) Findlay, J. A.; He, Z.-Q.; Calhoun, L. A. *J. Nat. Prod.* **1990**, 53, 1015; (c) Finamore, E.; Minale, L.; Riccio, R.; Rinaldo, G.; Zollo, F. *J. Org. Chem.* **1991**, 56, 1146; (d) Elliott, J. K.; Ross, D. M.; Pathirana, C.; Miao, S.; Andersen, R. J.; Singer, P. P.; Kokke, W. C. M. C.; Ayer, W. A. *Biol. Bull.* **1989**, 176, 73; (e) Burgoyne, D. L.; Miano, S.; Pathirana, C.; Andersen, R. J.; Ayer, W. A.; Singer, P. P.; Kokke, W. C. M. C.; Ross, D. M. *Can. J. Chem.* **1991**, 69, 20; (f) De Riccardis, F.; Minale, L.; Riccio, R.; Iorizzi, M.; Debitus, C.; Duhet, D.; Monniot, C. *Tetrahedron Lett.* **1993**, 34, 4381.
7. Terama, A.; De Ridder, C.; Kuenen, J. C.; Robertson, L. A. *Mar. Biol.* **1993**, 115, 179.
8. Bringmon, R. L.; De Pidder, C. *Appl. Environ. Microbiol.* **1998**, 64, 3491.